



Hypothesis

Anemia: A Potential Source of Bias in Clinical Trials of Angiogenesis Inhibitors: A Hypothesis



Reza Rastmanesh^{1,2,3*}

¹American Physical Society, College Park, MD, USA; ²The Nutrition Society, London, UK; ³Independent researcher, #6, Physicians Building, Sarshar Alley, Valis-Asr, Tajrish, Tehran, Iran

Received: February 16, 2023 | Revised: March 16, 2023 | Accepted: April 10, 2023 | Published online: May 11, 2023

Abstract

Although anemia may cause angiogenesis and neovascularization, especially in ocular situations, neither published nonrandomized clinical trials nor registered clinical trials have reported the anemia status as inclusion or exclusion criteria in their design. Increases in the circulating levels of erythropoietin and vascular endothelial growth factor are proportional to the levels of tissue hypoxia, which are influenced by hematocrit. Erythropoietin is a potent retinal angiogenic factor that is independent of endothelial growth factor and is capable of stimulating ischemia-induced retinal angiogenesis. We suggest that clinical trials investigating anti-vascular endothelial growth factor treatment for retinal neovascularization should measure appropriate variables such as serum erythropoietin, vascular endothelial growth factor, hemoglobin, and hematocrit to yield preliminary data for future trials of angiogenesis inhibitors. Ignoring the anemia status, serum erythropoietin, and/or vascular endothelial growth factor levels could create clinical uncertainty and subtle statistical bias in both systematic reviews and nonrandomized clinical trials that aim to evaluate the efficiency of angiogenesis inhibitors in several medical situations, including but not limited to ocular alterations, rheumatoid arthritis, and many types of cancer, just to mention a few. Implications of this type of bias could be involved in other disease situations in which angiogenesis inhibitors are used for medication, such as different carcinomas as well as metastases. In this hypothesis paper, we suggest that clinical trials of angiogenesis inhibitors should measure appropriate variables such as serum erythropoietin, hemoglobin, and hematocrit and match their participants by anemia and its severity to avoid a game-changing bias in their data analysis.

Introduction

Anemia and angiogenesis

Anemia is among the most common hematological manifestations of cancer, and it is estimated that almost 40–64% of patients treated for malignancies are affected by anemia. From a pathophysiological point of view, cancer-related anemia may be categorized into four broad but overlapping classes: hypoproliferative anemia comprising the common anemia of inflammation/chronic disease, miscellaneous etiologies, hemolytic anemia, and anemias

with uncertain etiologies. A positive connection is seen between tumor hypoxia and anemia. Hypoxemic situations enhance tumor growth and thereby resistance to medication through stimulating angiogenesis.¹ Erythropoietin (EPO) is considered as a hormone, a cytokine, and a growth factor. Its major function is regulation of erythropoiesis. EPO contributes to angiogenesis and neovascularization in the angiogenic switch of a tumor² and vascular endothelial growth factor (VEGF), which is a key dynamic molecule of angiogenesis.³ The development and application of anti-angiogenesis agents, specifically those targeting vascular VEGF, have become integral components of neovascularization therapies and anticancer regimens for many tumor types.

Anemia and angiogenesis inhibitors

The potential of angiogenesis inhibitors or angiogenesis-modulating agents as therapies for human diseases, namely eye diseases like retinal neovascularization and corneal neovascularization, has increased dramatically over the past 20 years due to newly emerged clinically available agents. Most of the recently published articles benefit from elegant and sophisticated methodology.⁴ However, although anemia is causally related to angiogenesis and neovas-

Keywords: Anemia; Bias; Clinical trials; Neovascularization; Angiogenesis inhibitor.
Abbreviations: Anti-VEGFR, anti-vascular endothelial growth factor receptor; EPO, erythropoietin; PDR, proliferative diabetic retinopathy; VEGF, vascular endothelial growth factor.
***Correspondence to:** Reza Rastmanesh, #6, Physicians Building, Sasha Alley, Vali Asr Street, Tajrish, Tehran 1961835555, Iran. ORCID: <https://orcid.org/0000-0002-6221-9062>. Tel: 98-2122750414, Fax: 98-2122735950, E-mail: r.rastmanesh@gmail.com/rezar@sbmu.ac.ir
How to cite this article: Rastmanesh R. Anemia: A Potential Source of Bias in Clinical Trials of Angiogenesis Inhibitors: A Hypothesis. *Explor Res Hypothesis Med* 2023;8(4):388–393. doi: 10.14218/ERHM.2023.00013.

cularization,⁵ especially in ocular situations,⁶ neither the published nonrandomized clinical trials nor the registered nonrandomized clinical trials have reported the anemia status as inclusion or exclusion criteria in their design. The main methodological issue here is whether the iron-deficiency status in enrolled patients confounds the efficacy of angiogenesis inhibitors, since a low hemoglobin level is associated with increased serum levels of vascular endothelial growth factor (VEGF) and it has been suggested that anemia might increase the progression of angiogenesis.⁷ Increases in the circulating levels of EPO are proportional to the levels of tissue hypoxia, which are influenced by hematocrit.⁸ In addition, there have been several reports that anemic patients have elevated levels of VEGF, which is a marker of tissue hypoxia.⁹ Moreover, it has been shown recently that the anemia of newborns induces EPO expression in the developing mouse retina.¹⁰

EPO and VEGF are independently associated with proliferative diabetic retinopathy (PDR), and EPO is more strongly associated with PDR than VEGF. EPO is a potent retinal angiogenic factor independent of VEGF and is capable of stimulating ischemia-induced retinal angiogenesis in PDR.¹¹ Additionally, the EPO-independent EPO-receptor signaling pathway has been demonstrated to play a potential role in cell proliferation and angiogenesis in human pterygium.¹² Furthermore, the gene expression levels of EPO and VEGF are upregulated in the murine ischemic retina, and the blockade of EPO inhibits retinal neovascularization *in vivo* and endothelial-cell proliferation in the vitreous of patients with PDR *in vitro*.¹³

Hypothesis

Through alteration of the endogenous EPO and VEGF levels, different types of anemia create a bias in trials of angiogenesis inhibitors. Interestingly, the results from an animal model of cancer showed that the antitumor efficacy of bevacizumab (a well-known anti-VEGF agent) was compromised in an anemic situation. Surprisingly, anti-vascular endothelial growth factor receptor (anti-VEGFR)-2, at the effective dose for normalization of systemic tissues and organs, did not show a significant antitumor effect in that cancer model. This was possibly because the plasma levels of EPO were significantly elevated in VEGF tumor-bearing mice. Also, VEGF-induced anemia was reversed by anti-VEGFR-2 but not by anti-VEGFR-1.¹⁴

These results may pertain to angiogenesis inhibitors and retinal neovascularization; in the human retina, VEGFR-1 neutralization decreased pigment epithelium-derived factor mRNA and protein expression, whereas anti-VEGFR-2 antibody had no effect.¹⁵ It should be kept in mind that a critical balance between VEGF and pigment epithelium-derived factor is important to prevent the development of retinal neovascularization.

Evaluation of hypothesis

If these findings can be extrapolated to human patients suffering from retinal neovascularization, a loss of anti-VEGF agent efficacy in patients suffering from iron deficiency or elevated plasma EPO levels might be involved. There is indirect evidence from human¹⁶ and direct evidence from animal^{14,17} studies that support our idea; for example, in patients with pre-existing higher EPO levels, the efficacy of anti-VEGF treatment was lower than that of patients with lower EPO levels.¹⁸ In addition, in an animal model of musculocutaneous tissue ischemia, flaps treated with EPO and bevacizumab did not show any change in tissue necrosis when

compared with animals receiving EPO only,¹⁹ supporting a possible counteraction between EPO and anti-VEGF agents. It might be argued that the interaction between locally administered exogenous anti-VEGF and endogenous intravitreal EPO and/or VEGF levels is complex. It should be kept in mind that there is a direct correlation between serum EPO and intravitreal EPO levels,²⁰ as well as between intravitreal EPO and intravitreal VEGF levels.²⁰ Very recently, it was found that the plasma VEGF level could predict the response in non-small cell lung cancer patients treated with bevacizumab.²¹ Interestingly, systemic dosing of squalamine lactate, an anti-VEGF agent, has yielded promising results in animal models as well as in humans.²²

Furthermore, there is strong evidence that changes in endogenous VEGF are related to the therapeutic effect of anti-VEGF agents.^{23,24} There are several reports of the high efficacy of systemic steroid treatment for retinal neovascularization, possibly through suppression of intravitreal VEGF. Overall, the results of these studies spark the hypothesis that the correlation between systemic and intravitreal EPO or VEGF levels is more plastic than previously anticipated. An anemic situation might have implications in anti-VEGF dose calculation as well. Such patients suffering from retinal neovascularization may need higher doses of anti-VEGF agents, while administering the same doses to nonanemic patients may increase the side effects of anti-VEGF agents. Further support for our claim comes from findings indicating increased anemia in exudative age-related macular degeneration.²⁵

Effects of different types of anemia and supplements on angiogenic/tumorigenic situations

It is noteworthy that different types of anemia and nutrient deficiencies leading to or contributing to anemia may have differential effects on the VEGF and EPO levels or their expression, and thereby on angiogenesis. Both severe and moderate iron deficiency^{5-7,26} as well as vitamin B12 deficiency²⁷⁻³³ lead to higher VEGF levels and angiogenesis, while the opposite is true in the case of folate deficiency anemia.³⁴⁻⁴² Other vitamins have their own mechanism for the regulation of angiogenesis.⁴³ Recently, Pedrosa and Lemes have reported a significantly enhanced gene expression of VEGF and HIF-1 α in patients with zinc deficiency and sickle cell anemia, and they showed an inverse dose-response association with hydroxyurea therapy.⁴⁴ Previously, higher serum VEGF levels in beta-thalassemia,⁴⁵⁻⁴⁸ hemolytic anemia,⁴⁹⁻⁵² idiopathic aplastic anemia⁵³⁻⁵⁵ and anemia of chronic disease⁵⁶⁻⁵⁸ have been well established. Etiologically, it is conceivable that different mechanisms are behind different types of anemia. Obviously, the interactions among the severity/types of anemia, nutrient reserves of the body, and the duration of the study/intervention can create an intricate and chaotic pattern, which is difficult to control by usual statistical methods.

There are many examples where the type of anemia and/or the type of nutrient supplementation can alter or modulate the process of tumorigenesis/angiogenesis as well as the therapy. For instance, some people take iron supplements without a medical indication. In a recent randomized controlled trial, it was shown that among comparable patients participating in age-related macular degeneration treatment trials, the use of iron supplements in participants undergoing bevacizumab treatment was correlated with retinal/subretinal hemorrhage in a dose-response fashion.⁵⁹ Ample literature exists on the deterioration and/or amelioration of tumorigenesis/angiogenesis following (un)indicated iron, folate, and vitamin B12 supplementation for different angiogenic/tumorigenic situations^{31,60-71} Figure 1 shows a simplified diagram of the

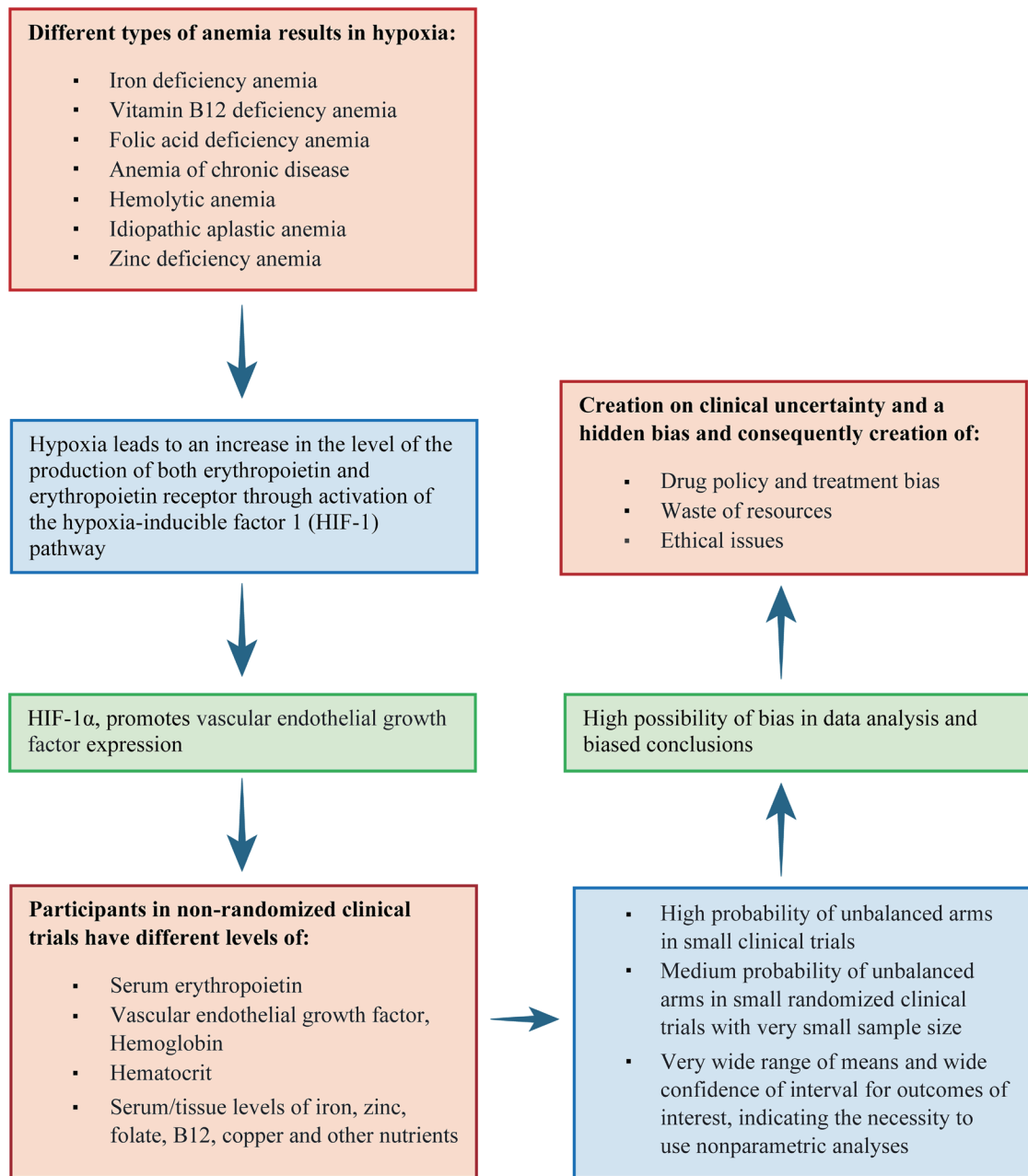


Fig. 1. A simplified diagram of mechanisms by which different types of anemia may create a bias in studies of angiogenesis inhibitors. HIF-1, hypoxia-inducible factor 1; HIF-1 α , hypoxia-inducible factor 1 alpha.

mechanisms by which different types of anemia may create a bias in studies of angiogenesis inhibitors.

Discussion

Ignoring the anemia status, serum EPO levels, and/or VEGF levels are likely examples of sources of bias in nonrandomized clinical trials that aim to evaluate the efficiency of angiogenesis inhibitors, especially in eye diseases. It might be argued that in clinical trials that randomly assign treatment or standard care to the study participants, the major advantage is that the treated and the un-

treated subjects are similar in many aspects because of the power of randomization, provided that a sufficient number of participants are enrolled (adequate power). It might also be argued that randomization is unlikely to lead to bias. However, a simple search of clinical trials registered in the National Institutes of Health database reveals that in almost half of the registered clinical trials, the design of the study is nonrandomized and uncontrolled.

Future directions

We would like to urge that all clinical trials on anti-VEGF agents

include the patient anemia status in their design. Obviously, the VEGF levels depend on the duration of anemia (chronic vs. acute), the definition of anemia, and the metabolic demand of the patients. Thus, it would be more important to measure the VEGF levels to meet the discussed problem more appropriately. To sufficiently randomize patients, data concerning the incidence of elevated EPO levels are necessary to plan a sufficient sample size. If researchers find it difficult to randomly allocate patients to the study arms or to balance the number of anemic/nonanemic patients in the study arms, like most other clinical trials, they can adjust the anemia status, preferably as the serum EPO level, as a potential confounder in the placebo and intervention groups when doing statistics, using proper statistical treatments.

One could argue that besides anemia, we should include genetic analysis, history of smoking, and food frequency questionnaires to measure anti-angiogenic food components into clinical trials. The point is that one should be able to estimate the size of the effect that these factors have based on scientific data prior to the planning of trials and to adjust the power calculation accordingly. The goal is to evenly distribute these factors between the treatment arm(s) and control arm(s) of a randomized controlled trial. So, if it is not possible to correct for all of these factors, the concept of randomization comes into play. Due to the high impact of confounding factors in clinical trials investigating angiogenesis inhibitors, the results of nonrandomized clinical trials might lead to a major bias in reviews and guideline protocols. On the other hand, in randomized clinical trials, the power of the study has to be so strong that all of these confounding factors can be equally distributed between the groups.

Conclusions

We suggest that the significance of systematic reviews could be enhanced by including all factors and giving an estimation of the effect size of the affected group. This could positively impact the design of contemporary clinical trials. Implications of this kind of bias could be involved in other disease situations in which angiogenesis inhibitors/promoters are used for medication, such as different carcinomas as well as metastases. Implications of this kind of bias also could be involved both in situations of higher endogenous EPO production and exogenous EPO administration, just to mention a few examples.

Acknowledgments

None.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of interest

The author declares no conflict of interest.

References

- [1] Gaspar BL, Sharma P, Das R. Anemia in malignancies: pathogenetic and diagnostic considerations. *Hematology* 2015;20(1):18–25. doi:10.1179/1607845414Y.0000000161, PMID:24666207.
- [2] Annese T, Tamma R, Ruggieri S, Ribatti D. Erythropoietin in tumor angiogenesis. *Exp Cell Res* 2019;374(2):266–273. doi:10.1016/j.yexcr.2018.12.013, PMID:30576679.
- [3] Goswami AG, Basu S, Huda F, Pant J, Ghosh Kar A, *et al.* An appraisal of vascular endothelial growth factor (VEGF): the dynamic molecule of wound healing and its current clinical applications. *Growth Factors* 2022;40(3-4):73–88. doi:10.1080/08977194.2022.2074843, PMID:35584274.
- [4] Tanioka H, Shimada K, Tsuji A, Kochi M, Kim HM, Takahashi T, *et al.* Phase II Study of S-1 and Irinotecan Plus Bevacizumab as Second-line Treatment for Patients With Metastatic Colorectal Cancer Resistant to the Fluoropyrimidine-oxaliplatin-cetuximab Regimen. *Anticancer Res* 2022;42(5):2675–2681. doi:10.21873/anticancer.15745, PMID:35489740.
- [5] Zohora F, Bidad K, Pourpak Z, Moin M. Biological and immunological aspects of iron deficiency anemia in cancer development: A narrative review. *Nutr Cancer* 2018;70(4):546–556. doi:10.1080/01635581.2018.1460685, PMID:29697284.
- [6] Rastmanesh R. Possibility of enhanced risk of retinal neovascularization in repeated blood donors: blood donation and retinal alteration. *Int J Gen Med* 2011;4:647–656. doi:10.2147/IJGM.S23206, PMID:21941450.
- [7] Kang HS, Shin AY, Yeo CD, Park CK, Kim JS, Kim JW, *et al.* Clinical significance of anemia as a prognostic factor in non-small cell lung cancer carcinoma with activating epidermal growth factor receptor mutations. *J Thorac Dis* 2020;12(5):1895–1902. doi:10.21037/jtd-19-3932, PMID:32642093.
- [8] Chang X, Li Q, Tang H. Use of preoperative erythropoietin therapy to facilitate autologous blood donation in orthopedic surgery: A meta-analysis. *Medicine (Baltimore)* 2020;99(2):e18577. doi:10.1097/MD.00000000000018577, PMID:31914036.
- [9] Howarth C, Banerjee J, Eaton S, Aladangady N. Biomarkers of gut injury in neonates - where are we in predicting necrotising enterocolitis? *Front Pediatr* 2022;10:1048322. doi:10.3389/fped.2022.1048322, PMID:36518779.
- [10] Scheerer N, Dünker N, Imagawa S, Yamamoto M, Suzuki N, Fandrey J. The anemia of the newborn induces erythropoietin expression in the developing mouse retina. *Am J Physiol Regul Integr Comp Physiol* 2010;299(1):R111–118. doi:10.1152/ajpregu.00108.2010, PMID:20463184.
- [11] Loukovaara S, Sandholm J, Aalto K, Liukkonen J, Jalkanen S, Yegutkin GG. Deregulation of ocular nucleotide homeostasis in patients with diabetic retinopathy. *J Mol Med (Berl)* 2017;95(2):193–204. doi:10.1007/s00109-016-1472-6, PMID:27638339.
- [12] Kase S, Kitaichi N, Furudate N, Yoshida K, Ohno S. Increased expression of mucinous glycoprotein KL-6 in human pterygium. *Br J Ophthalmol* 2006;90(9):1208–1209. doi:10.1136/bjo.2006.094300, PMID:16929070.
- [13] Watanabe D, Suzuma K, Matsui S, Kurimoto M, Kiryu J, Kita M, *et al.* Erythropoietin as a retinal angiogenic factor in proliferative diabetic retinopathy. *N Engl J Med* 2005;353(8):782–792. doi:10.1056/NEJMoa041773, PMID:16120858.
- [14] Farzam P, Johansson J, Mireles M, Jiménez-Valerio G, Martínez-Lozano M, Choe R, *et al.* Pre-clinical longitudinal monitoring of hemodynamic response to anti-vascular chemotherapy by hybrid diffuse optics. *Biomed Opt Express* 2017;8(5):2563–2582. doi:10.1364/BOE.8.002563, PMID:28663891.
- [15] Bhattarai N, Hytti M, Reinisalo M, Kaarniranta K, Mysore Y, Kauppinen A. Hydroquinone predisposes for retinal pigment epithelial (RPE) cell degeneration in inflammatory conditions. *Immunol Res* 2022;70(5):678–687. doi:10.1007/s12026-022-09300-0, PMID:35661979.
- [16] Vroeling L, Lind JS, de Haas RR, Verheul HM, van Hinsbergh VW, Broxterman HJ, *et al.* CD133+ circulating haematopoietic progenitor cells predict for response to sorafenib plus erlotinib in non-small cell lung cancer patients. *Br J Cancer* 2010;102(2):268–275. doi:10.1038/sj.bjc.6605477, PMID:20010948.
- [17] Xue Y, Religa P, Cao R, Hansen AJ, Lucchini F, Jones B, *et al.* Anti-VEGF agents confer survival advantages to tumor-bearing mice by improving cancer-associated systemic syndrome. *Proc Natl Acad Sci USA* 2008;105(47):18513–18518. doi:10.1073/pnas.0807967105, PMID:19017793.
- [18] Rezaeian F, Wettstein R, Egger JF, Sandmann F, Rucker M, Tobalem M,

- et al.* Erythropoietin-induced upregulation of endothelial nitric oxide synthase but not vascular endothelial growth factor prevents musculo-cutaneous tissue from ischemic damage. *Lab Invest* 2010;90(1):40–51. doi:10.1038/labinvest.2009.117, PMID:19901910.
- [19] Wang B, Zhang X, Chen H, Koh A, Zhao C, Chen Y. A Review of Intraocular Biomolecules in Retinal Vein Occlusion: Toward Potential Biomarkers for Companion Diagnostics. *Front Pharmacol* 2022;13:859951. doi:10.3389/fphar.2022.859951, PMID:35559255.
- [20] Stahl A, Buchwald A, Martin G, Junker B, Chen J, Hansen LL, *et al.* Vitreal levels of erythropoietin are increased in patients with retinal vein occlusion and correlate with vitreal VEGF and the extent of macular edema. *Retina* 2010;30(9):1524–1529. doi:10.1097/IAE.0b013e3181d37539, PMID:20664492.
- [21] Li B, Jiang C, Xu Y, Fan X, Yang L, Zou B, *et al.* Genome-wide DNA methylation signature predict clinical benefit of bevacizumab in non-small cell lung cancer. *BMC Cancer* 2022;22(1):828. doi:10.1186/s12885-022-09918-1, PMID:35906610.
- [22] Mammari N, Salles E, Beaussart A, El-Kirat-Chatel S, Varbanov M. Squalamine and its aminosterol derivatives: Overview of biological effects and mechanisms of action of compounds with multiple therapeutic applications. *Microorganisms* 2022;10(6):1205. doi:10.3390/microorganisms10061205, PMID:35744723.
- [23] Bockhorn M, Goralski M, Prokofiev D, Dammann P, Grünewald P, Trippler M, *et al.* VEGF is important for early liver regeneration after partial hepatectomy. *J Surg Res* 2007;138(2):291–299. doi:10.1016/j.jss.2006.07.027, PMID:17275844.
- [24] Tsuchihashi S, Ke B, Kaldas F, Flynn E, Busuttill RW, Briscoe DM, *et al.* Vascular endothelial growth factor antagonist modulates leukocyte trafficking and protects mouse livers against ischemia/reperfusion injury. *Am J Pathol* 2006;168(2):695–705. doi:10.2353/ajpath.2006.050759, PMID:16436682.
- [25] Naito Y, Tsujino T, Matsumoto M, Sakoda T, Ohyanagi M, Masuyama T. Adaptive response of the heart to long-term anemia induced by iron deficiency. *Am J Physiol Heart Circ Physiol* 2009;296(3):H585–593. doi:10.1152/ajpheart.00463.2008, PMID:19136608.
- [26] Devi S, Mukhopadhyay A, Dwarkanath P, Thomas T, Crasta J, Thomas A, *et al.* Combined vitamin B-12 and balanced protein-energy supplementation affect homocysteine remethylation in the methionine cycle in pregnant south Indian women of low vitamin B-12 status. *J Nutr* 2017;147(6):1094–1103. doi:10.3945/jn.116.241042, PMID:28446631.
- [27] Lewerenz J, Gocht A, Hoeger PH, von den Driesch P, Eckert B, Lamszus K, *et al.* Multiple vascular abnormalities and a paradoxical combination of vitamin B12 deficiency and thrombocytosis in a case with POEMS syndrome. *J Neurol* 2003;250(12):1488–1491. doi:10.1007/s00415-003-0261-7, PMID:14673584.
- [28] Mani C, Kochhar P, Ravikumar G, Dwarkanath P, Sheela CN, George S, *et al.* Placental expression of ENG, VEGF, and FLT: Gender-specific associations with maternal vitamin B₁₂ status. *Eur J Clin Nutr* 2020;74(1):176–182. doi:10.1038/s41430-019-0449-2, PMID:31209272.
- [29] Rathod R, Khaire A, Kale A, Joshi S. A combined supplementation of vitamin B₁₂ and n-3 polyunsaturated fatty acids across two generations improves nerve growth factor and vascular endothelial growth factor levels in the rat hippocampus. *Neuroscience* 2016;339:376–384. doi:10.1016/j.neuroscience.2016.10.018, PMID:27743986.
- [30] Rathod RS, Khaire AA, Kale AA, Joshi SR. Maternal omega-3 fatty acid supplementation to a vitamin B12 deficient diet normalizes angiogenic markers in the pup brain at birth. *Int J Dev Neurosci* 2015;43:43–49. doi:10.1016/j.ijdevneu.2015.04.006, PMID:25889224.
- [31] Reddy SS, Prabhakar YK, Kumar CU, Reddy PY, Reddy GB. Effect of vitamin B₁₂ supplementation on retinal lesions in diabetic rats. *Mol Vis* 2020;26:311–325. PMID:32355441.
- [32] Vacca A, Ria R, Ribatti D, Semeraro F, Djonov V, Di Raimondo F, *et al.* A paracrine loop in the vascular endothelial growth factor pathway triggers tumor angiogenesis and growth in multiple myeloma. *Haematologica* 2003;88(2):176–185. PMID:12604407.
- [33] Bendardaf R, Sharif-Askari FS, Sharif-Askari NS, Syrjänen K, Pyrhönen S. Patients with hMLH1 or/and hMSH2-deficient Metastatic Colorectal Cancer Are Associated with Reduced Levels of Vascular Endothelial Growth Factor-1 Expression and Higher Response Rate to Irinotecan-based Regimen. *Anticancer Res* 2018;38(11):6399–6404. doi:10.21873/anticancer.13000, PMID:30396964.
- [34] Davis CK, Nampoothiri SS, Rajanikant GK. Folic Acid Exerts Post-Ischemic Neuroprotection In Vitro Through HIF-1 α Stabilization. *Mol Neurobiol* 2018;55(11):8328–8345. doi:10.1007/s12035-018-0982-3, PMID:29542054.
- [35] Hyodo T, Ito Y, Hosono K, Uematsu S, Akira S, Majima M, *et al.* The role of mPGES-1 in promoting granulation tissue angiogenesis through regulatory T-cell accumulation. *In Vivo* 2022;36(5):2061–2073. doi:10.21873/invivo.12932, PMID:36099134.
- [36] Iloki-Assanga S, McCarty MF. Nutraceutical targeting of placental synthesis of soluble Fms-like tyrosine Kinase-1 (sFlt-1) as strategy for preventing and controlling pre-eclampsia. *Curr Pharm Des* 2018;24(20):2255–2263. doi:10.2174/1381612824666180723162327, PMID:30039754.
- [37] Li Y, Gao R, Liu X, Chen X, Liao X, Geng Y, *et al.* Folate Deficiency could restrain decidual angiogenesis in pregnant mice. *Nutrients* 2015;7(8):6425–6445. doi:10.3390/nu7085284, PMID:26247969.
- [38] Mathur RS, Mathur SP. In vitro downregulation of growth factors by insulin-like growth factor binding protein-3 in cervical cancer. *Gynecol Onco* 2003;91(2):410–415. doi:10.1016/s0090-8258(03)00513-4, PMID:14599874.
- [39] Tuska RM, Helm SM, Graf CF, James C, Kong G, Stiemsma LT, *et al.* Surfeit folic acid, protein, and exercise modify oncogenic inflammatory biomarkers and fecal microbiota. *Front Nutr* 2023;9:1060212. doi:10.3389/fnut.2022.1060212, PMID:36742002.
- [40] Westin SN, Herzog TJ, Coleman RL. Investigational agents in development for the treatment of ovarian cancer. *Invest New Drugs* 2013;31(1):213–229. doi:10.1007/s10637-012-9837-3, PMID:22661305.
- [41] Yin J, Yi J, Yang C, Xu B, Lin J, Hu H, *et al.* Weiqi Decoction attenuated chronic atrophic gastritis with precancerous lesion through regulating microcirculation disturbance and HIF-1 α signaling pathway. *Evid Based Complement Alternat Med* 2019;2019:2651037. doi:10.1155/2019/2651037, PMID:31320912.
- [42] Saghiri MA, Asatourian A, Ershadifar S, Moghadam MM, Sheibani N. Vitamins and regulation of angiogenesis: [A, B1, B2, B3, B6, B9, B12, C, D, E, K]. *J Func Foods* 2017;38:180–196.
- [43] Pedrosa AM, Lemes RPG. Gene expression of HIF-1 α and VEGF in response to hypoxia in sickle cell anaemia: Influence of hydroxycarbamide. *Br J Haematol* 2020;190(1):e39–e42. doi:10.1111/bjh.16693, PMID:32352161.
- [44] Åström M, Hahn-Strömberg V, Zetterberg E, Vedin I, Merup M, Palmblad J. X-linked thrombocytopenia with thalassemia displays bone marrow reticulin fibrosis and enhanced angiogenesis: comparisons with primary myelofibrosis. *Am J Hematol* 2015;90(3):E44–48. doi:10.1002/ajh.23907, PMID:25421114.
- [45] Fahmey SS, Naguib HF, Abdelshafy SS, Alashry RE. Vascular endothelial growth factor in children with thalassemia major. *Mediterr J Hematol Infect Dis* 2013;5(1):e2013044. doi:10.4084/MJHID.2013.044, PMID:23795282.
- [46] Olgar S, Kara A, Hicyilmaz H, Balta N, Canatan D. Evaluation of angiogenesis with vascular endothelial growth factor in patients with thalassemia major. *Pediatr Int* 2010;52(2):247–251. doi:10.1111/j.1442-200X.2009.02956.x, PMID:19744226.
- [47] Tantawy AAG, Adly AAM, Ismail EAR, Youssef OI, Ali ME. Soluble fms-like tyrosine kinase 1 as a link between angiogenesis and endothelial dysfunction in pediatric patients with β -thalassemia intermedia. *Clin Appl Thromb Hemost* 2017;23(8):943–950. doi:10.1177/1076029617692879, PMID:28301910.
- [48] Carlucci S, Stabile G, Catagini S, Borghi C, Scutiero G, Morano D, *et al.* Fetal disseminated intravascular coagulopathy, hydrops and massive umbilical vein thrombosis consequence of a rare placental condition: multifocal chorangiomas. *J Matern Fetal Neonatal Med* 2022;35(20):4009–4013. doi:10.1080/14767058.2020.1843154, PMID:33143492.
- [49] Négrier S, Pérol D, Bahleda R, Hollebecque A, Chatelut E, Boyle H, *et al.* Phase I dose-escalation study of pazopanib combined with bevacizumab in patients with metastatic renal cell carcinoma or other advanced tumors. *BMC Cancer* 2017;17(1):547. doi:10.1186/s12885-017-3527-7, PMID:28810837.
- [50] Pellé G, Shweke N, Duong Van Huyen JP, Tricot L, Hessaïne S,

- Frémeaux-Bacchi V, *et al.* Systemic and kidney toxicity of intraocular administration of vascular endothelial growth factor inhibitors. *Am J Kidney Dis* 2011;57(5):756–759. doi:10.1053/j.ajkd.2010.11.030, PMID:21295897.
- [51] Yonem O, Arslan S, Sokmensuer C, Salmanzade S. Can angiogenesis be a target of treatment for ribavirin associated hemolytic anemia? *Hepato-gastroenterology* 2010;57(99-100):562–566. PMID:20698227.
- [52] Deng S, Zeng Y, Wu L, Hu Z, Shen J, Shen Y, *et al.* The regulatory roles of VEGF-Notch signaling pathway on aplastic anemia with kidney deficiency and blood stasis. *J Cell Biochem* 2019;120(2):2078–2089. doi:10.1002/jcb.27516, PMID:30230583.
- [53] Gupta P, Khurana N, Singh T, Gupta D, Dhingra KK. Bone marrow angiogenesis in aplastic anemia—a study of CD 34 and VEGF expression in bone marrow biopsies. *Hematology* 2009;14(1):16–21. doi:10.1179/102453309X385070, PMID:19154660.
- [54] Li JP, Zheng CL, Han ZC. Abnormal immunity and stem/progenitor cells in acquired aplastic anemia. *Crit Rev Oncol Hematol* 2010;75(2):79–93. doi:10.1016/j.critrevonc.2009.12.001, PMID:20045349.
- [55] Čubranić A, Dobrila-Dintinjana R, Redžović A, Dintinjana M, Petranović D, Golčić M. Endogenous erythropoietin and erythropoietin receptors in colorectal cancer; can we answer the questions? *Med Hypotheses* 2016;96:16–19. doi:10.1016/j.mehy.2016.09.017, PMID:27959268.
- [56] Besarab A, Hemmerich S. Anemia of Chronic Disease. In: Provenzano R, Lerma EV, Szczec L (eds). *Management of Anemia: A Comprehensive Guide for Clinicians*. New York: Springer; 2018:43–80. doi:10.1007/978-1-4939-7360-6.
- [57] Muchnik E, Kaplan J. HIF prolyl hydroxylase inhibitors for anemia. *Expert Opin Investig Drugs* 2011;20(5):645–656. doi:10.1517/13543784.4.2011.566861, PMID:21406036.
- [58] Song D, Ying GS, Dunaief JL, Bhuyan R, Li Y, Maguire MG, *et al.* Association between oral iron supplementation and retinal or subretinal hemorrhage in the comparison of age-related macular degeneration treatment trials. *Retina* 2019;39(10):1965–1972. doi:10.1097/IAE.0000000000002295, PMID:30157115.
- [59] Abri Aghdam K, Soltan Sanjari M, Ghasemi Falavarjani K. Erythropoietin in ophthalmology: A literature review. *J Curr Ophthalmol* 2016;28(1):5–11. doi:10.1016/j.joco.2016.01.008, PMID:27239595.
- [60] Blazovics A. Small molecules in cancer therapy: Cytotoxics and molecularly targeted agents. *Curr Signal Transd Therapy* 2011;6(1):2–19.
- [61] Cartron PF, Hervouet E, Debien E, Olivier C, Poulliquen D, Menanteau J, *et al.* Folate supplementation limits the tumorigenesis in rodent models of gliomagenesis. *Eur J Cancer* 2012;48(15):2431–2441. doi:10.1016/j.ejca.2012.01.002, PMID:22325970.
- [62] Chai SM, Mathur R, Ong SG. Retinal vasculopathy in Fanconi anemia. *Ophthalmic Surg Lasers Imaging* 2009;40(5):498–500. doi:10.3928/15428877-20090901-11, PMID:19772276.
- [63] Ciappio ED, Liu Z, Brooks RS, Mason JB, Bronson RT, Crott JW. Maternal B vitamin supplementation from preconception through weaning suppresses intestinal tumorigenesis in *Apc1638N* mouse offspring. *Gut* 2011;60(12):1695–1702. doi:10.1136/gut.2011.240291, PMID:21659408.
- [64] Gerald D, Berra E, Frapart YM, Chan DA, Giaccia AJ, Mansuy D, *et al.* JunD reduces tumor angiogenesis by protecting cells from oxidative stress. *Cell* 2004;118(6):781–794. doi:10.1016/j.cell.2004.08.025, PMID:15369676.
- [65] Kim YI. Role of folate in colon cancer development and progression. *J Nutr* 2003;133(11):3731S–3739S. doi:10.1093/jn/133.11.3731S, PMID:14608107.
- [66] Pascual M, Bohle B, Alonso S, Mayol X, Salvans S, Grande L, *et al.* Preoperative administration of erythropoietin stimulates tumor recurrence after surgical excision of colon cancer in mice by a vascular endothelial growth factor-independent mechanism. *J Surg Res* 2013;183(1):270–277. doi:10.1016/j.jss.2012.12.041, PMID:23348072.
- [67] Rahat B, Mahajan A, Bagga R, Hamid A, Kaur J. Epigenetic modifications at DMRs of placental genes are subjected to variations in normal gestation, pathological conditions and folate supplementation. *Sci Rep* 2017;7:40774. doi:10.1038/srep40774, PMID:28098215.
- [68] Sasaki K, Duan J, Murohara T, Ikeda H, Shintani S, Shimada T, *et al.* Rescue of hypercholesterolemia-related impairment of angiogenesis by oral folate supplementation. *J Am Coll Cardiol* 2003;42(2):364–72. doi:10.1016/s0735-1097(03)00629-6, PMID:12875777.
- [69] Souza E, Cho KH, Harris ST, Flindt NR, Watt RK, Pai AB. Hypoxia-inducible factor prolyl hydroxylase inhibitors: a paradigm shift for treatment of anemia in chronic kidney disease? *Expert Opin Investig Drugs* 2020;29(8):831–844. doi:10.1080/13543784.2020.1777276, PMID:32476498.
- [70] Suk KK, Dunbar JA, Liu A, Daher NS, Leng CK, Leng JK, *et al.* Human recombinant erythropoietin and the incidence of retinopathy of prematurity: a multiple regression model. *J AAPOS* 2008;12(3):233–238. doi:10.1016/j.jaapos.2007.08.009, PMID:18589385.
- [71] van Patot MC, Gassmann M. Hypoxia: adapting to high altitude by mutating EPAS-1, the gene encoding HIF-2 α . *High Alt Med Biol* 2011;12(2):157–167. doi:10.1089/ham.2010.1099, PMID:21718164.